

# Pathogenesis, diagnosis and prevention of dysbiosis in broilers

IWT - 135067

Ghent University, Faculty of Veterinary Medicine

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Department of Reproduction, Obstetrics, and Herd Health,  
Epidemiology Unit

15th January 2018

# Challenges to the poultry industry that resulted in the project

Wet litter  
Dysbiosis



Footpad lesions



Antimicrobial usage  
and AMR

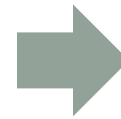


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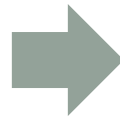


# Challenges to the poultry industry that resulted in the project

Wet litter  
Dysbiosis

Footpad lesions

Antimicrobial usage  
and AMR



Intestinal  
Health



# Strategic aims on long term

- Improve animal welfare
- Increase sustainability of poultry industry
- Prevent disease
- Diagnose gut health problems

# Specific aims of the project

What is intestinal health?  
What is dysbiosis?

What are factors that  
influence dysbiosis?

What are easy-to-measure  
markers for dysbiosis?

How can we prevent dysbiosis?

# Workpackages

- WP1. Identification of risk factors in the field
- WP2. Description of microbiota composition and histology in field dysbiosis cases
- WP3. Development of diagnostic tools
- WP4. Model development and validation of preventive measures

# Pathogenesis, diagnosis and prevention of dysbiosis in broilers

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Development of an *in vivo* model for dysbiosis in broilers (WP4)

→ macroscopic lesion scoring parameters

→ morphological changes of the intestinal wall

→ intestinal inflammatory parameters

} WP2

→ markers for intestinal wall integrity

→ microbial marker populations

} WP3

# WP4

## Validation of diagnostic criteria

### Aims

- Task 1) Development *in vivo* model dysbiosis
- Task 2) In field validation of prevention protocols

# In vivo dysbiosis model: set-up

- Control group

D1	D12	D12 - D18	D19 - D21	D20
Starter mash	57% wheat + 5% rye			

- Challenge group

D1	D12	D12 - D18	D19 - D21	D20
Starter mash	57% wheat + 5% rye	Antibiotics	<b>Bacterial challenge</b> <ul style="list-style-type: none"><li>- <i>E. coli</i></li><li>- <i>C. perfringens</i></li></ul>	<b>Coccidiosis challenge</b> <ul style="list-style-type: none"><li>- <i>Eimeria acervulina</i></li><li>- <i>Eimeria maxima</i></li></ul>

## Sampling

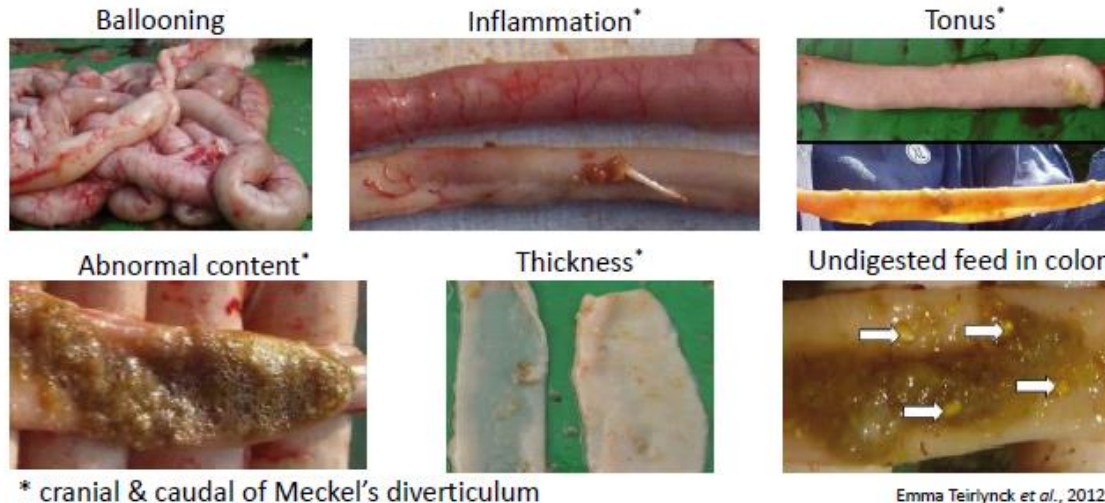
- D26 & D28
- 2 treatments, 9 replicates, 3 animals/group = 108 animals

# Macroscopic lesion scoring of the intestine to identify dysbiosis

1) Body weight (BW)

2) Dysbiosis score (DS)

→ sum of 10 macroscopic parameters scored 0 (absent) or 1 (present)

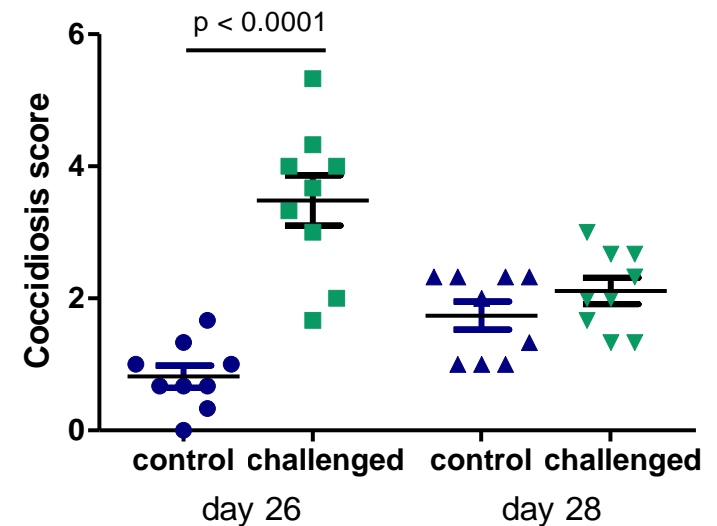
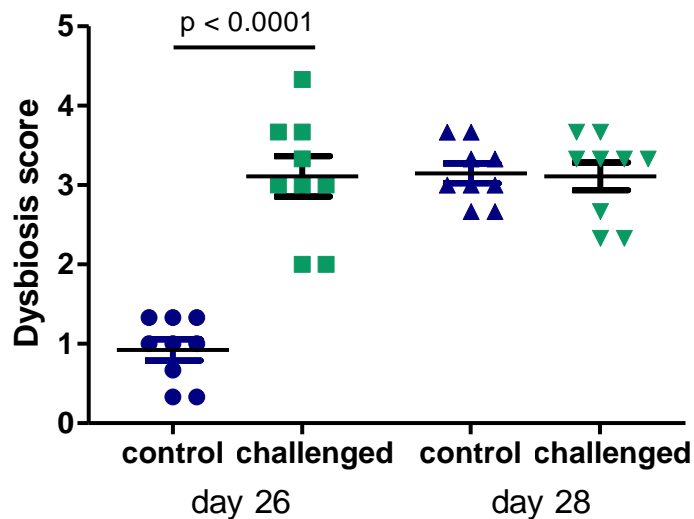
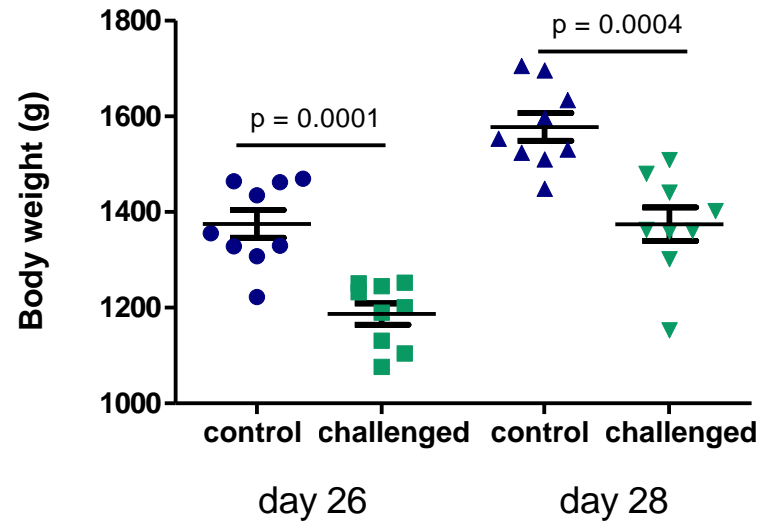


3) Coccidiosis score (CS) (Tellez et al., 2014)

- *E. acervulina*
  - *E. maxima*
  - *E. tenella*
- scored 0 (absent) to 4 (severe) → total score = sum

# Macroscopic scoring parameters

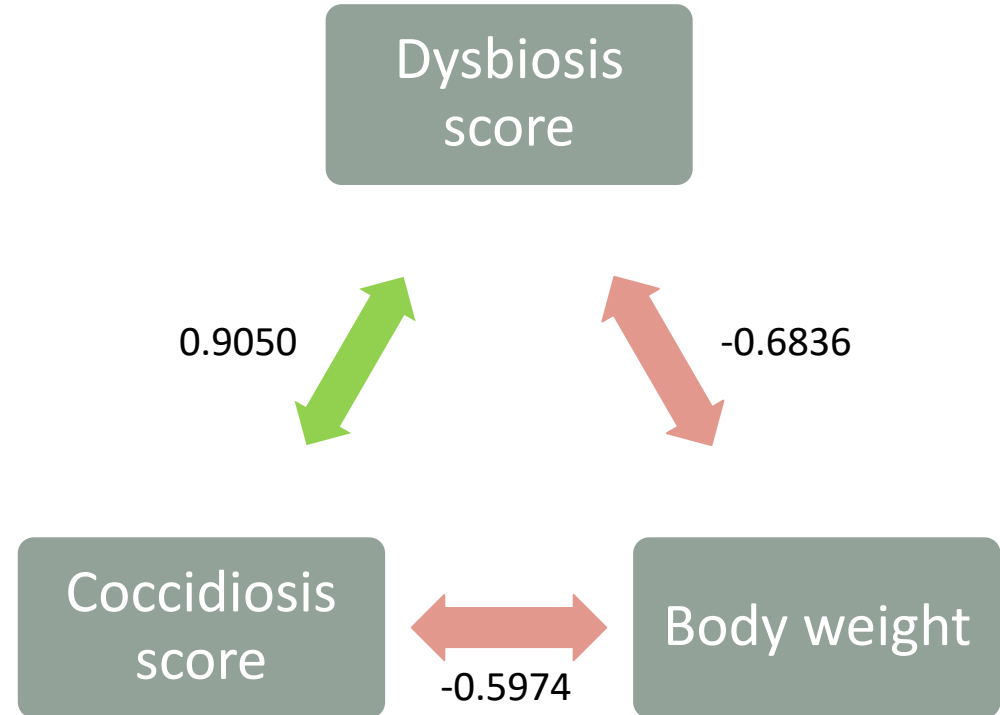
→ All animals



# Macroscopic scoring parameters

## D26 – associations

- Positive (0 to 1)
- Negative (0 to -1)



## D28

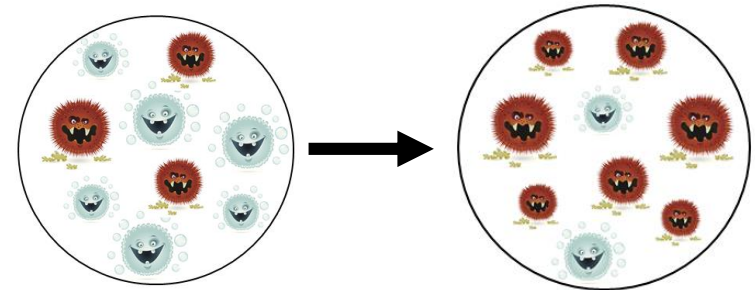
→ no significant associations

# WP2

**Description of morphological  
characteristics & gastrointestinal  
inflammatory parameters**

## Determination dysbiosis

- Definition still vague
- Macroscopic lesion scoring system = only method
- 1 study: scoring system  $\leftrightarrow$  morphological parameters



## Aims

- Task 1) Description of morphological changes of the intestinal wall
- Task 2) Description of intestinal inflammatory parameters



# Description morphological changes

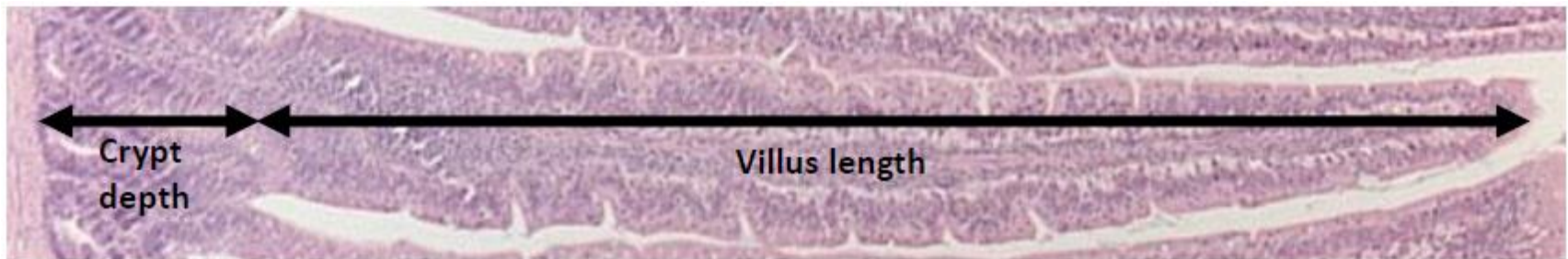
Duodenum (all animals)



Haematoxylin & eosin staining



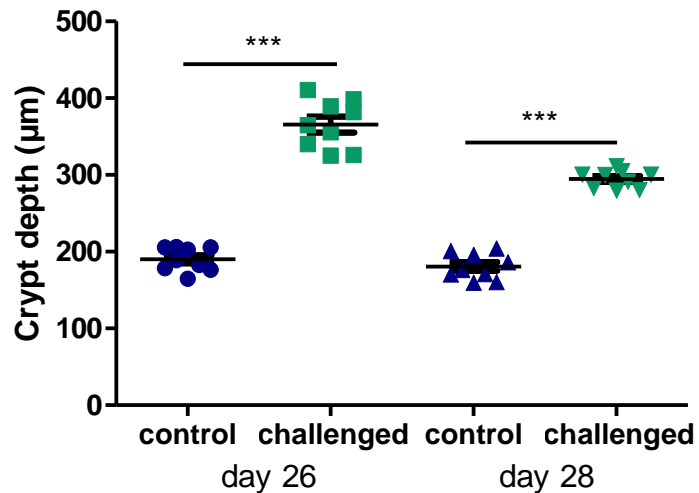
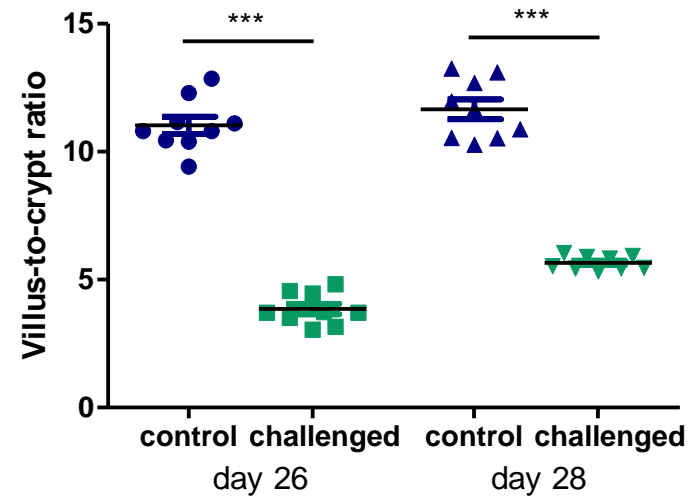
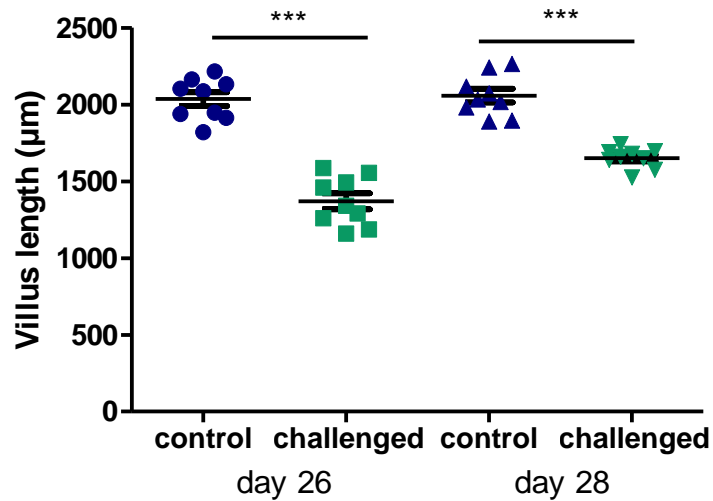
Measurement using computer based image analysis program



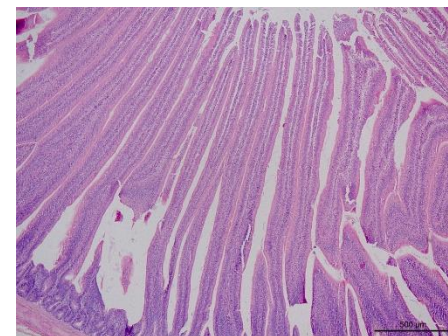
Histological parameters

- villus length
  - crypt depth
- } Villus-to-crypt ratio

# Description morphological changes



\*\*\*  $p < 0.0001$



Control



Challenged

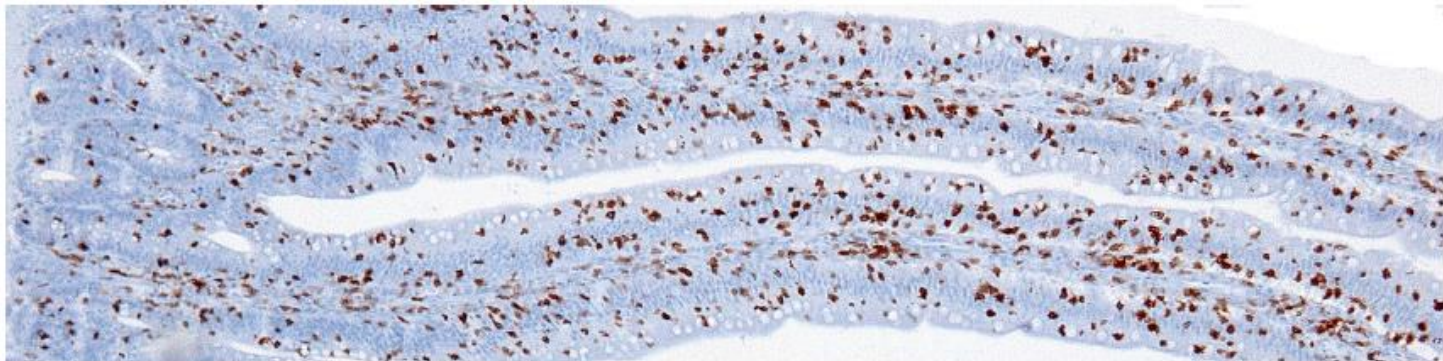
Duodenum (all animals)



Immunohistochemistry (CD<sub>3</sub> staining)

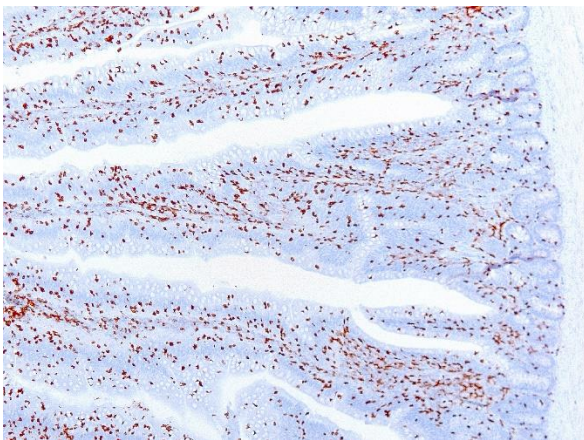
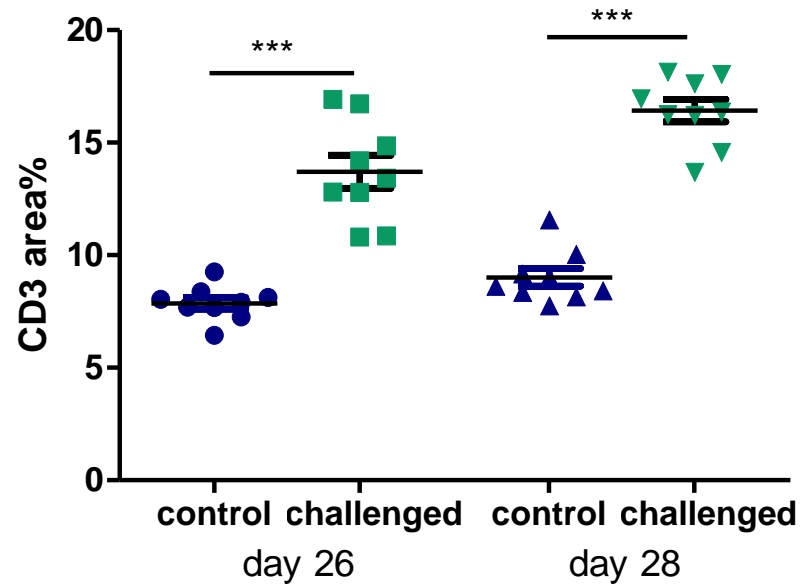


Computer based image analysis system



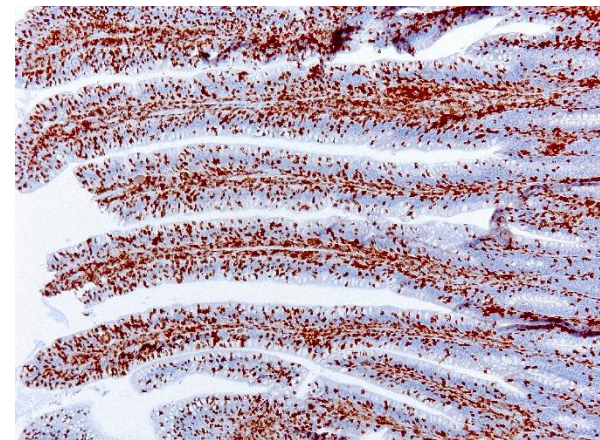
Measurement area% of CD<sub>3</sub><sup>+</sup> T-cells  
→ quantification infiltration immune cells

# Description inflammatory parameters



Control

\*\*\*  $p < 0.0001$

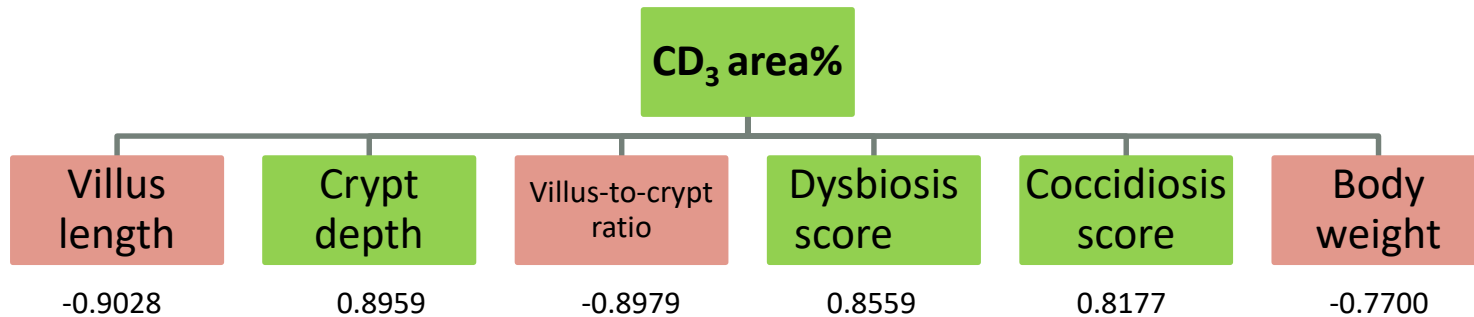
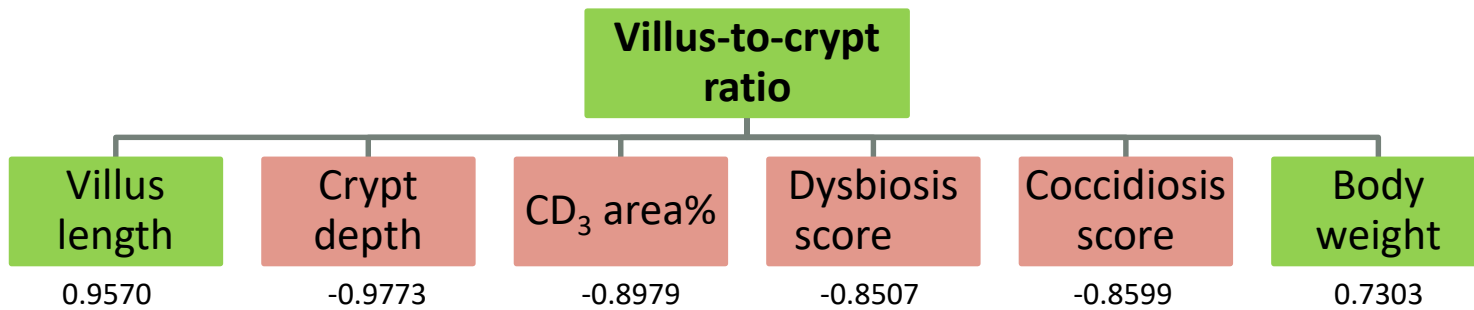


Challenged

# Description morphological changes & inflammatory parameters

## Associations D26

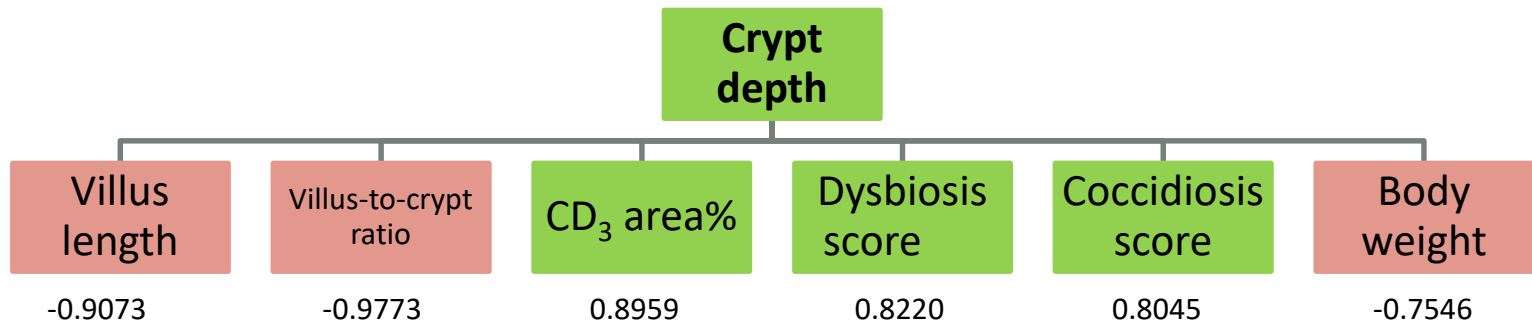
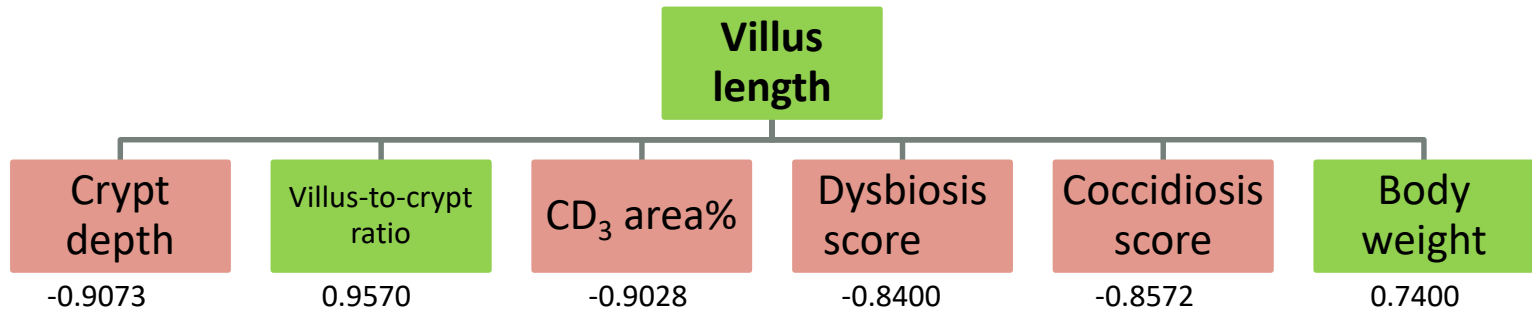
Positive: p1 ↑ then p2 ↑  
Negative: p1 ↑ then p2 ↓



# Description morphological changes & inflammatory parameters

## Associations D26

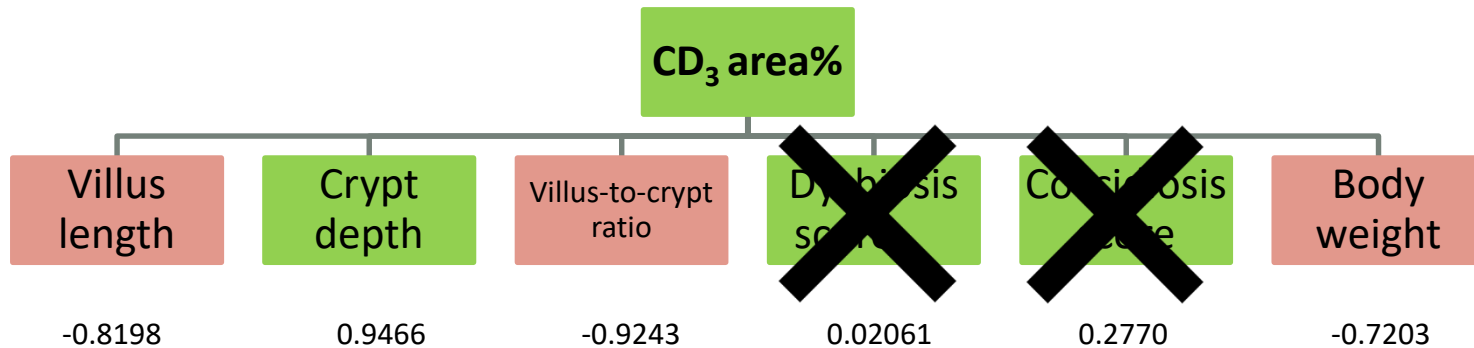
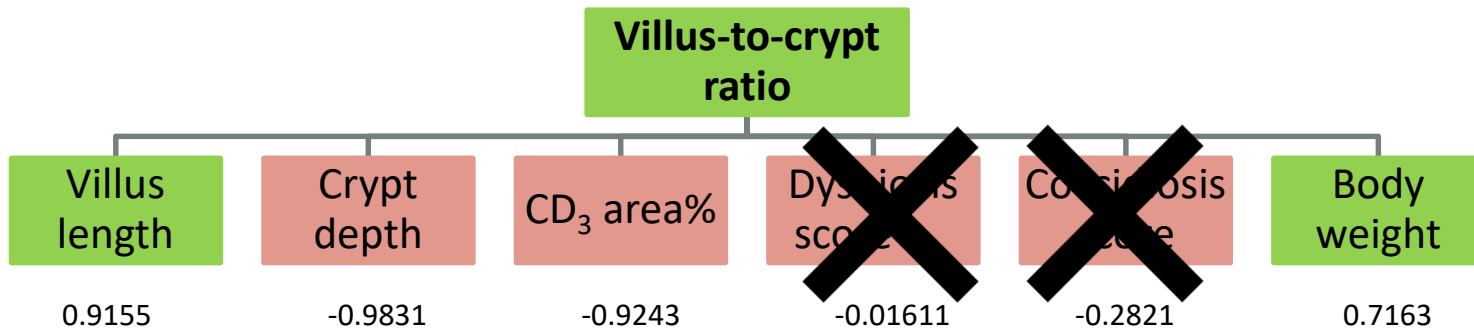
Positive: p1 ↑ then p2 ↑  
Negative: p1 ↑ then p2 ↓



# Description morphological changes & inflammatory parameters

## Associations D28

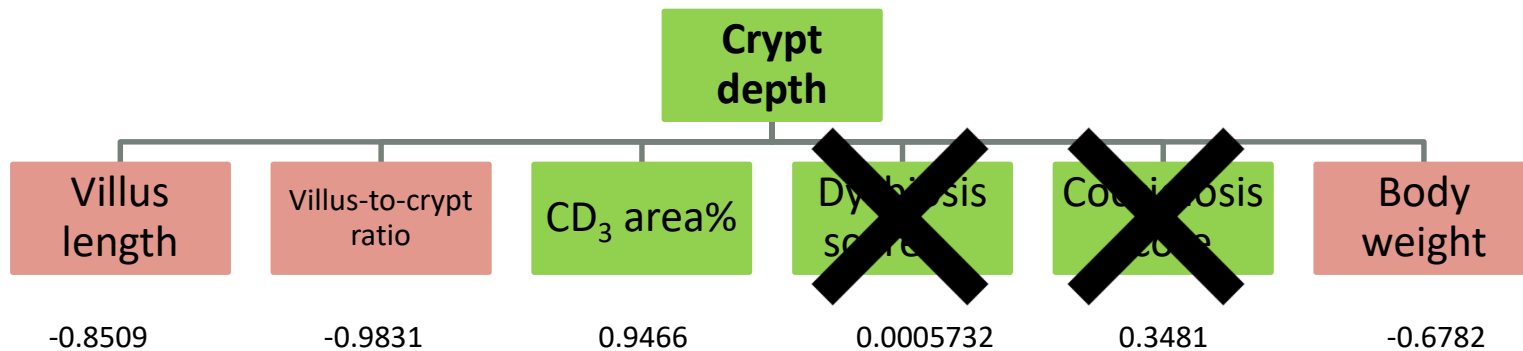
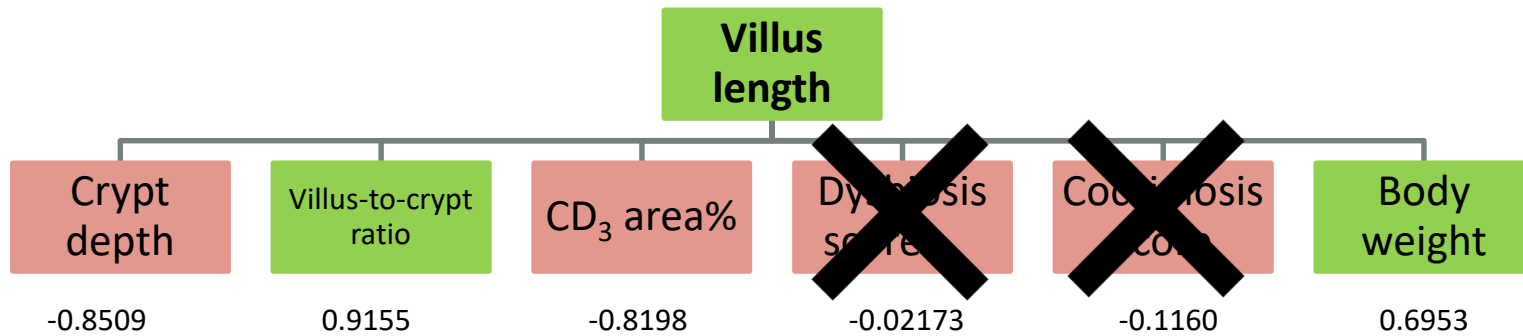
Positive: p1 ↑ then p2 ↑  
Negative: p1 ↑ then p2 ↓



# Description morphological changes & inflammatory parameters

## Associations D28

Positive: p1 ↑ then p2 ↑  
Negative: p1 ↑ then p2 ↓





Dysbiosis score dependent of coccidiosis score

→ not identical

Day 26

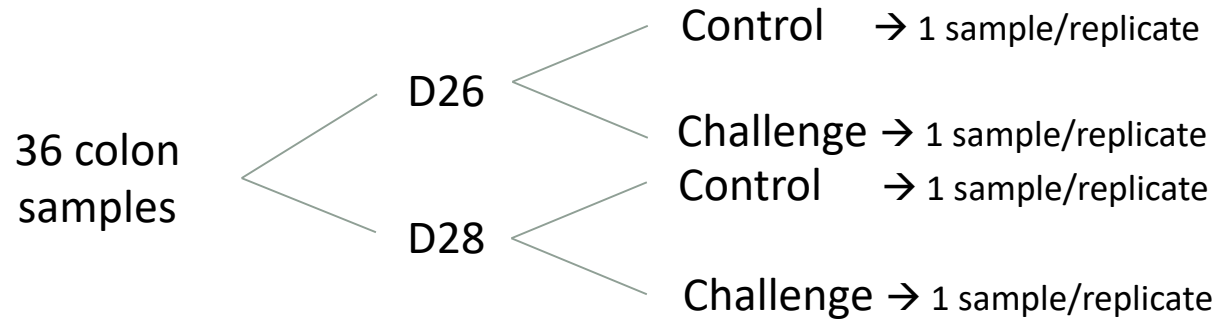
→ Association between all macroscopic and histological parameters

Day 28

→ No more association with dysbiosis & coccidiosis score

## **WP3**

# **Development of diagnostic criteria for nonspecific gastrointestinal problems in broilers**



## Discovery proteomics

Liquid chromatography–mass spectrometry (LC-MS)



## Analysis with Progenesis

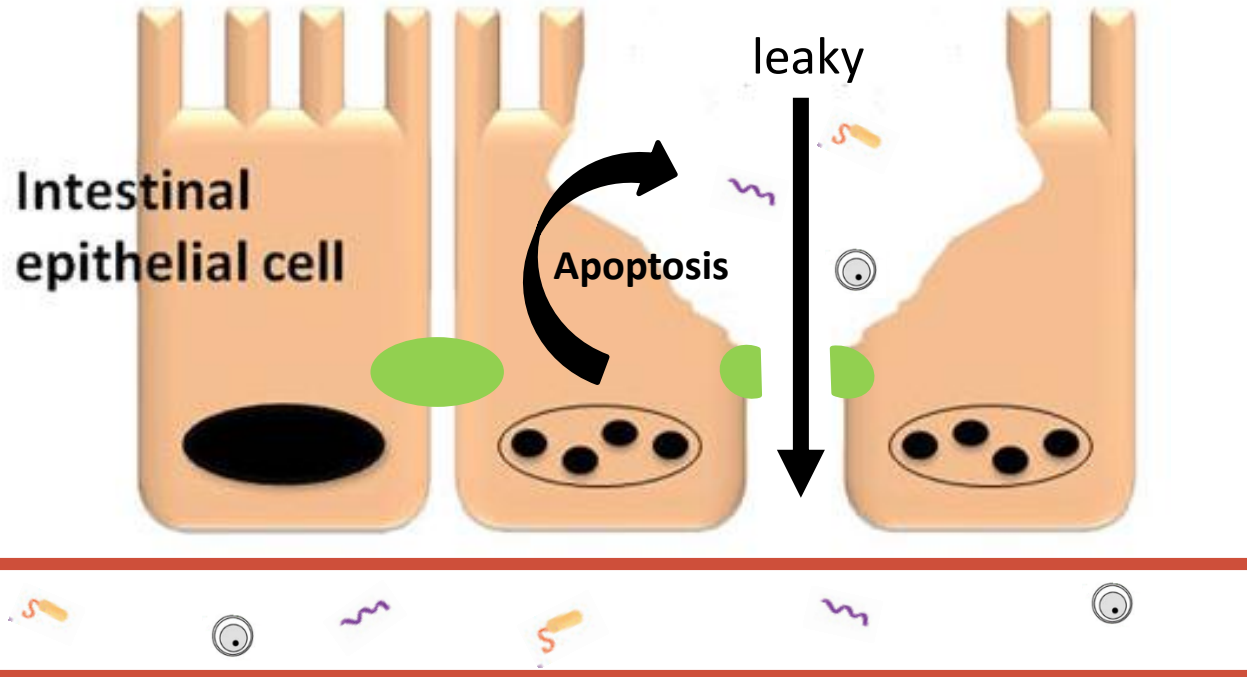


Checked associations between proteins and

- villus length
- crypt depth
- villus-to-crypt ratio
- CD<sub>3</sub> area%
- dysbiosis score

# Proteomics

Lumen



# Proteomics

Inflammation marker	Villus length	CD <sub>3</sub>	Dysbiosis	Function
Myeloid protein 1	-	+	+	granulocyt
Tissue destruction markers	Villus length	CD <sub>3</sub>	Dysbiosis	Funtion
Fibronectin	-	+	+	Component of extracellular matrix
Annexin A5	-			Calcium-channel activity
Nucleophosmin	-	+		Activates tumor suppressor gene p53
Epithelial cell markers	Villus length	CD <sub>3</sub>	Dysbiosis	Function
Carbonic anhydrase 2	-		+	Alkaline secretion
Aminopeptidase Ey	+	-	-	Secreted for protein digestion
Leakage markers	Villus length	CD <sub>3</sub>	Dysbiosis	Function
Transthyretin	-	+		Acute phase protein
Ovotransferrin	-			Acute phase protein
Ovoinhibitor	-			Serine protease inhibitor
Apolipoprotein A1	-			Component of high-density lipoprotein
Tight junction marker	Villus length	CD <sub>3</sub>	Dysbiosis	Function
Alpha-actinin	-		+	Actin-binding protein

# Ovotransferrin ELISA

Colon content (all animals)



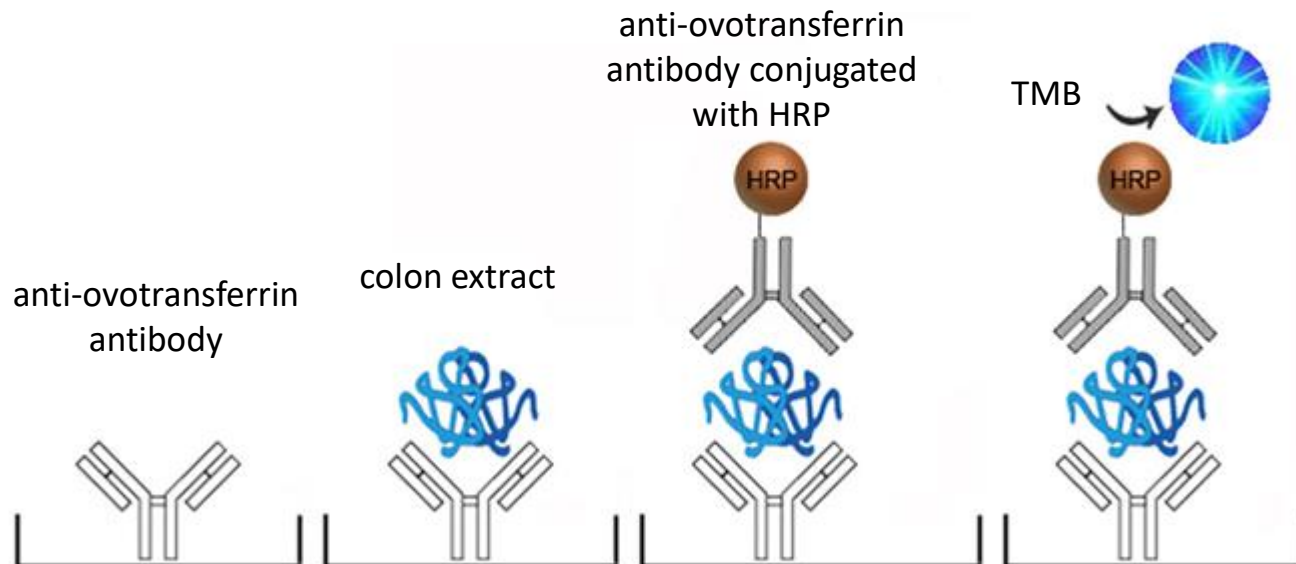
Extraction in PBS



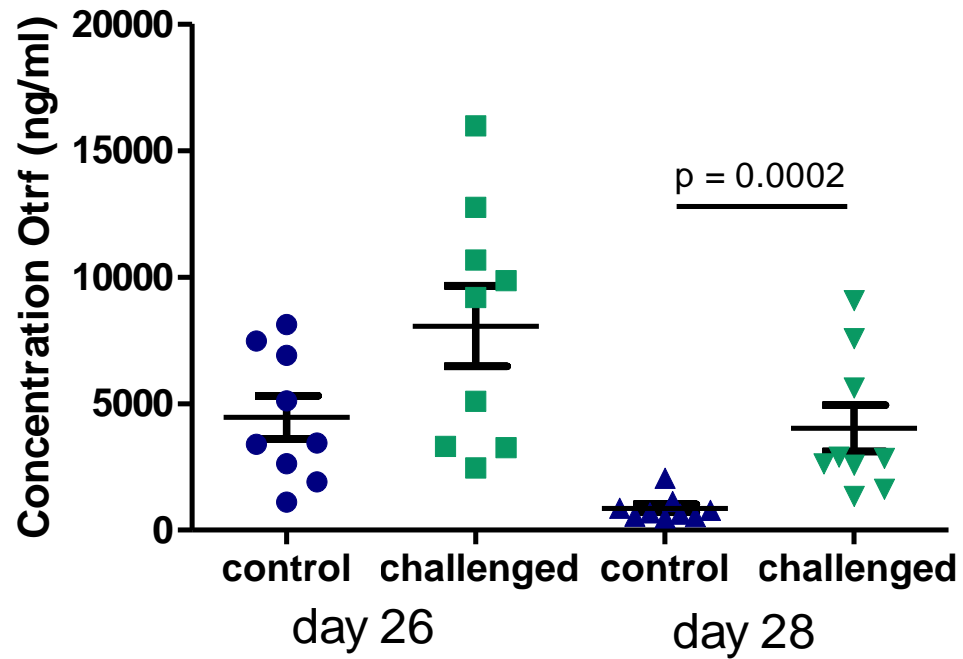
Double antibody sandwich ELISA



Concentration determination ovotransferrin

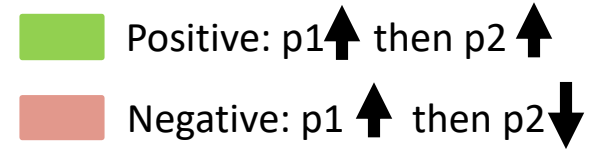


# Ovotransferrin ELISA



# Ovotransferrin ELISA

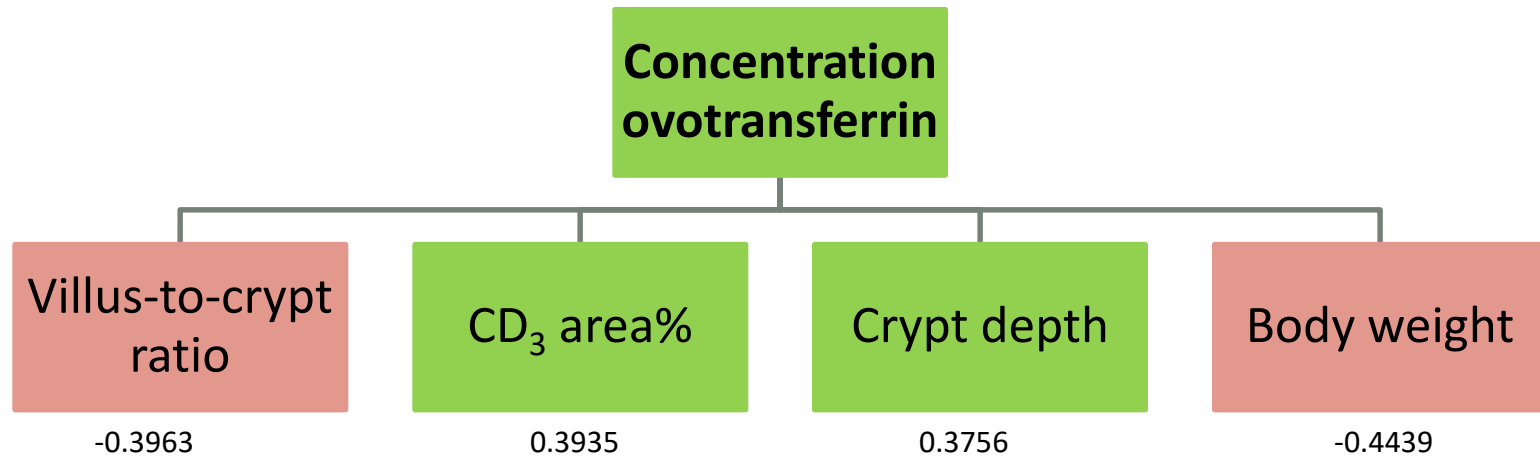
## Associations



### D26



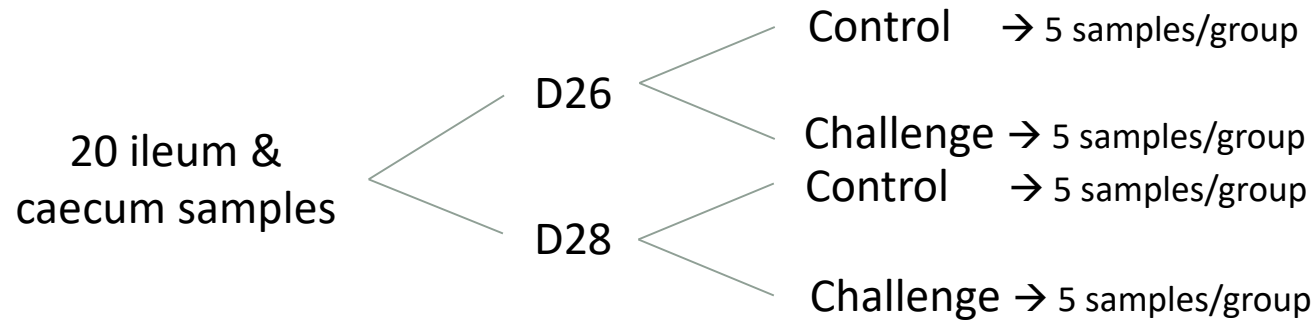
### D28





- D26 > protein expression
- Identification of protein markers
  - fibronectin
  - alpha-actinin
  - myeloid protein 1
  - carbonic anhydrase 2
  - aminopeptidase
- ➔ Description of gut health
  - intestinal damage
  - inflammation

# 16S sequencing



16S sequencing via Illumina MiSeq technology



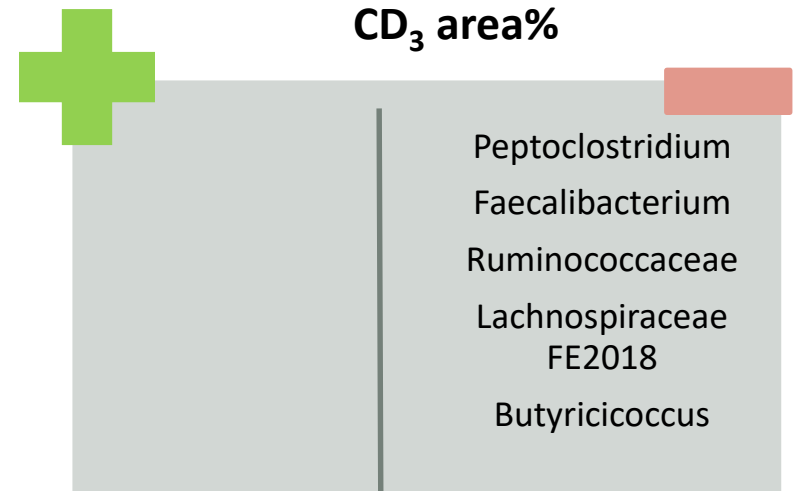
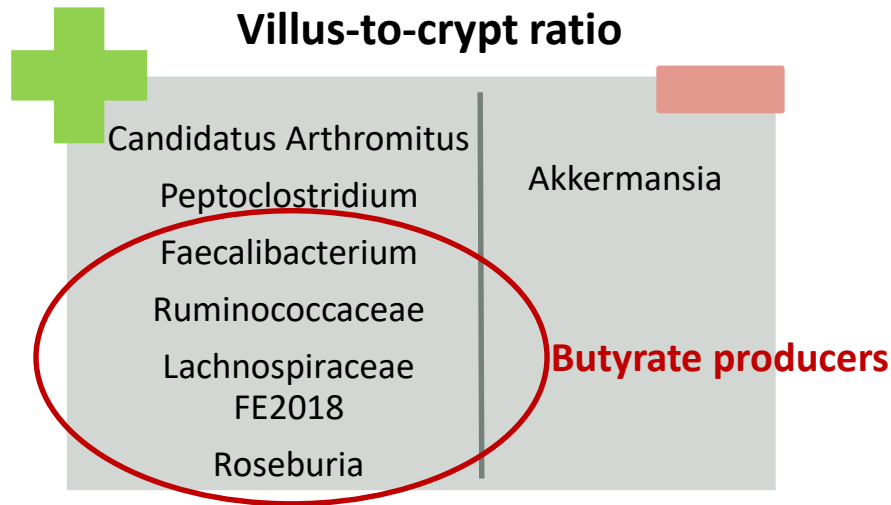
Checked associations between 16s data and

- macroscopic parameters
- histology parameters

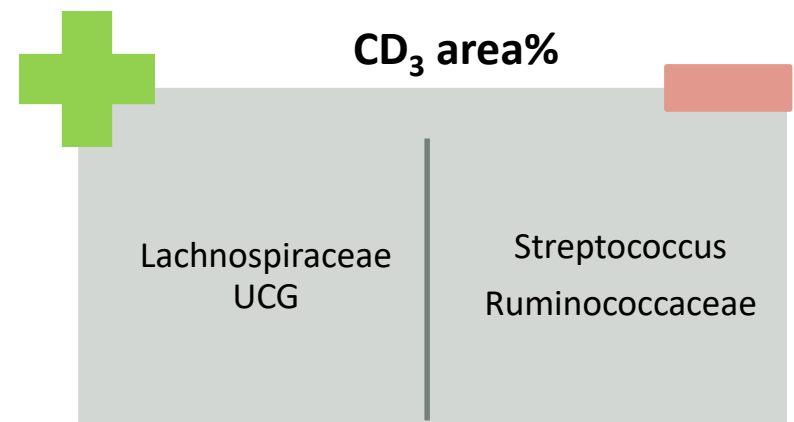
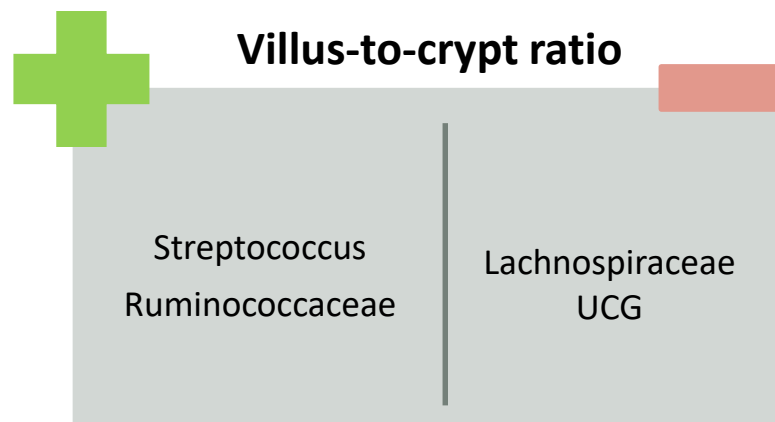
# 16S sequencing

## Associations – caecum

D26



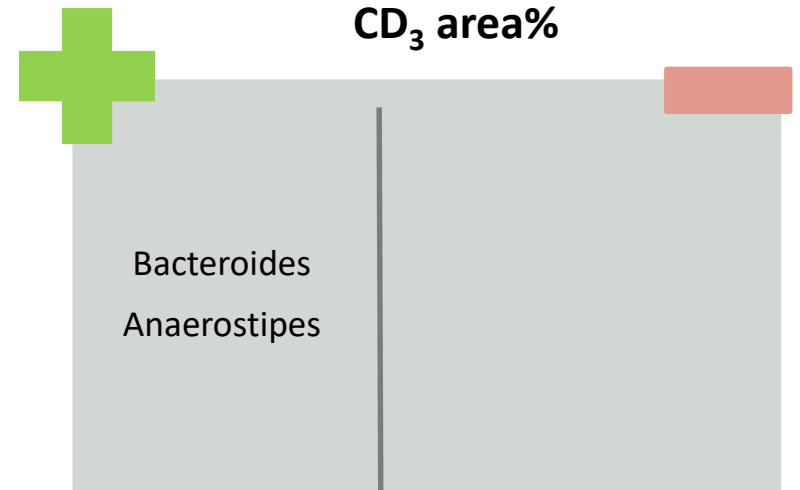
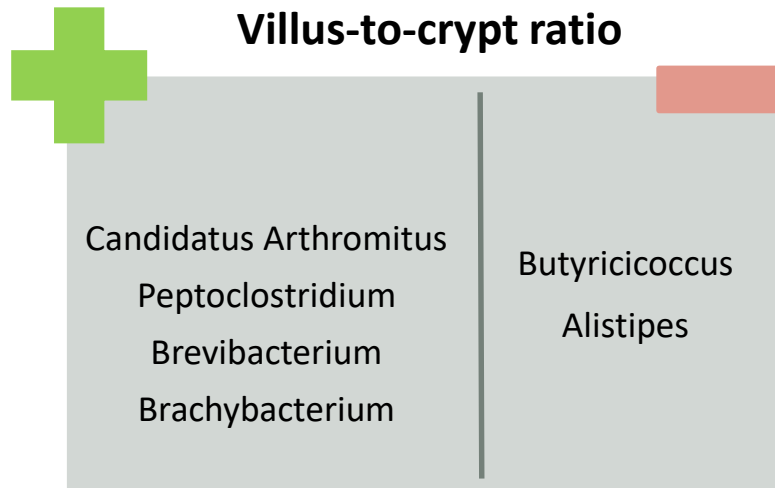
D28



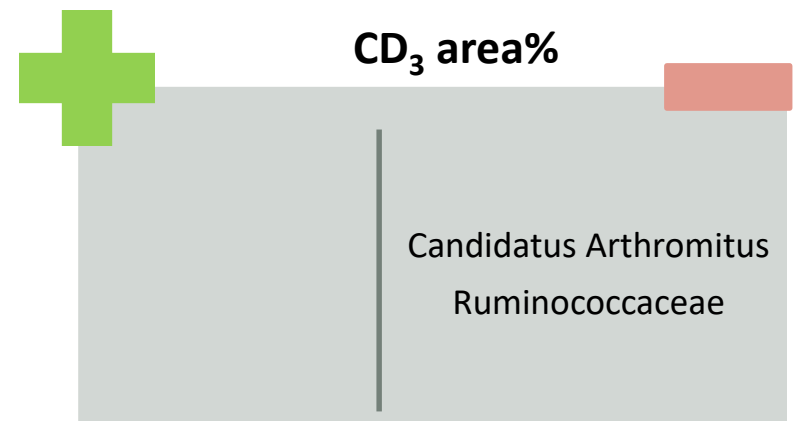
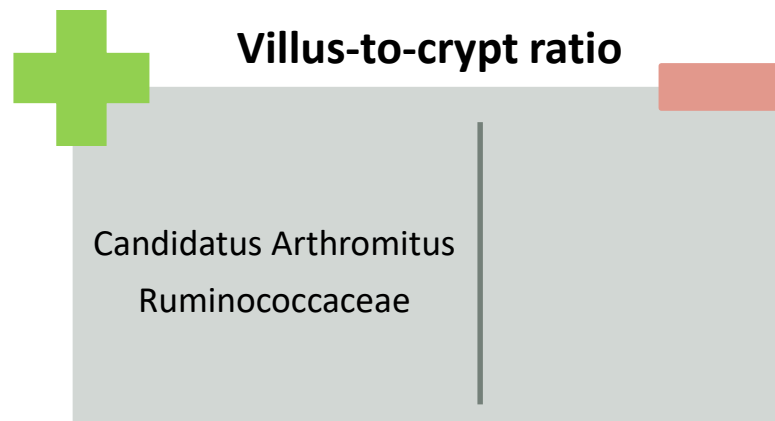
# 16S sequencing

## Associations – ileum

D26



D28



Ileum & caecal samples (1 replicate/group at D26 & D28)

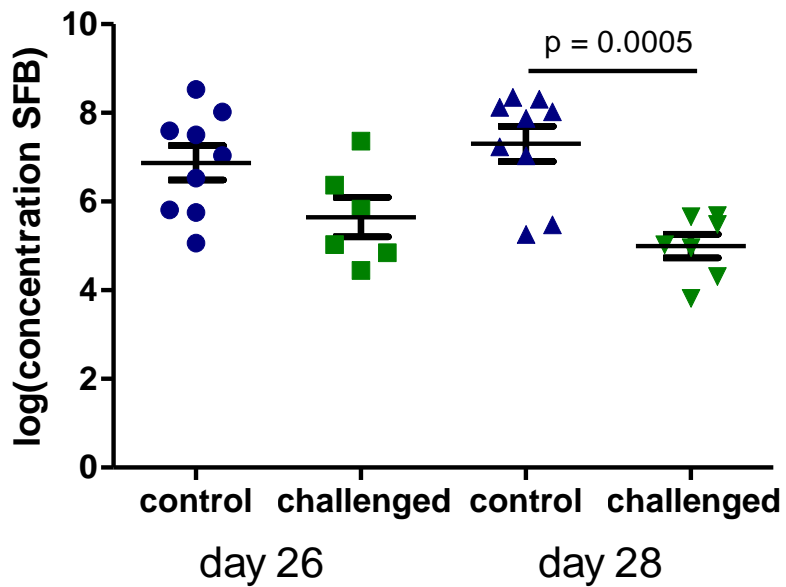


Quantification with qPCR  
Segmented filamentous bacteria (Candidatus Arthromitus)

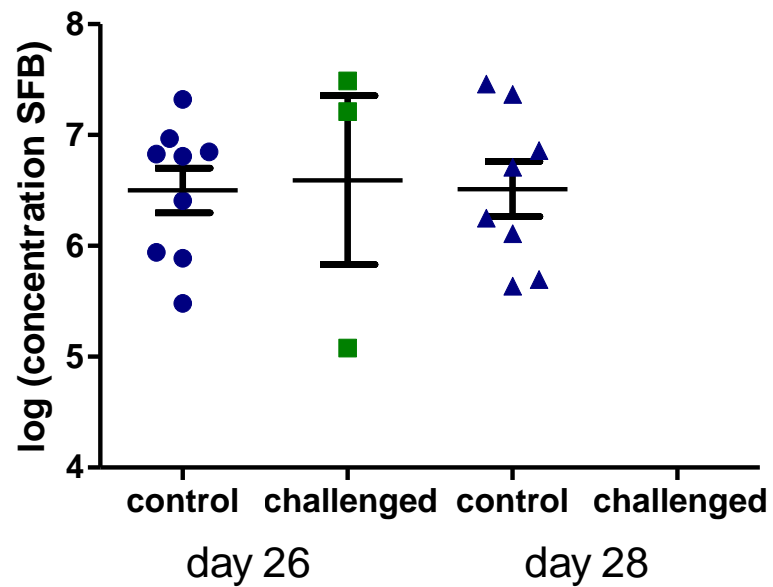


Check with 16s data

## Ileum



## Caecum

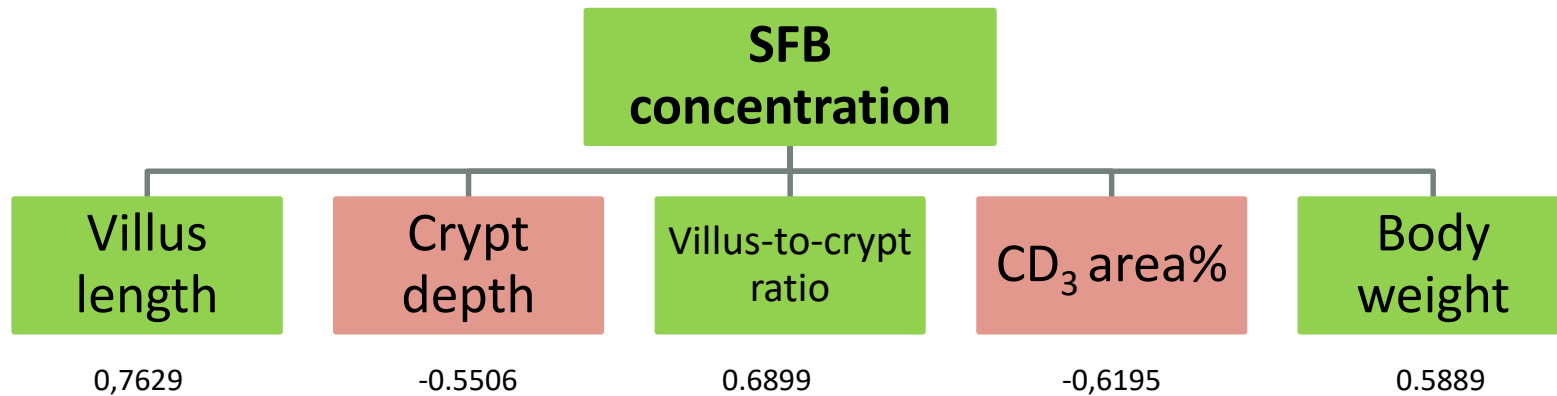


# qPCR

D26

→ No associations

D28



→ Same observations as 16S data

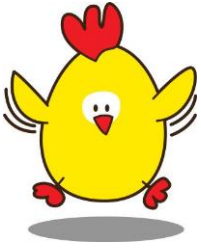
## D26

- Association between macroscopic and histology parameters
- > protein expression
- > microbial diversity

➔ Model for evaluation of gut health



- Validation model
- Analysis field samples
- Proteomics of ileal samples
- ELISA development for quantification protein markers
- qPCR microbial data



**Thanks for your attention!**